

Gut microbiome metagenomic sequences of honey bees (*Apis mellifera*) exposed to crops

L. Tran,¹ L. Lansing,² M. Cunningham,^{1,3} J. Ho,¹ T. Deckers,¹ T. Newman,¹ L. Wu,¹ A. S. Gregoris,¹ J. Zorz,² L. Muntz,¹ K. Lee,¹ D. Trépanier-Leroux,¹ I. M. Conflitti,⁴ M. Pepinelli,⁴ E. M. Walsh,¹ N. Morfin,⁵ J. E. Powell,⁶ N. Moran,⁶ S. E. Hoover,⁷ S. F. Pernal,¹ R. W. Currie,⁸ P. Giovenazzo,⁹ E. Guzman-Novoa,⁵ H. Jabbari,³ L. J. Foster,¹⁰ A. Zayed,⁴ R. Ortega Polo,² M. M. Guarna^{1,3}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT The gut microbiome of the European honey bee (*Apis mellifera*) is vital to its health, yet large-scale studies are scarce. We present metagenomic sequencing data from 180 samples collected near and far from eight crops across Canada over 2 years. These data sets will help address various biological and environmental questions.

KEYWORDS honey bees, metagenomics, gut microbiome

Microbiomes play a crucial role in agroecosystems as well as in individual animal and plant health (1, 2). European honey bees (*Apis mellifera*) rely heavily on their gut microbiome for nutrition, health, and immunity (3). However, large-scale studies are still scarce. We announce the release of shotgun metagenomic sequencing data from 180 honey bee samples collected near and far from eight different crops (apple, canola oil, canola seed, corn, cranberry, highbush blueberry, lowbush blueberry, and soybean) across Canada in 2020 and 2021 (Table 1). Four honey bee colonies were placed at five sites near to, or far from, each crop as previously described (4, 5). We generally defined “near” as within 0.5 km of the crop and “far” as greater than 1.5 km from the crop.

For highbush blueberry, bees were collected at four time points: before (time point 1), during (time point 2), at the end of (time point 3), and after (time point 4) the pollination period (4). For the other crops, bees for microbiome analyses were collected only at time point 2. Bee samples were processed by isolating whole guts, i.e., digestive tracts excluding the honey crop. Each replicate consisted of pooled guts from 12 individual bees (i.e., three bees from each of the four colonies located at each site). After homogenization, samples were shipped on dry ice to the Centre d'expertise et des services Génome Québec/Génome Québec Centre of Expertise and Services (Montréal, Quebec, Canada) for genomic DNA extraction and quantification (Quant-iT PicoGreen dsDNA Assay Kit, Life Technologies), library construction (NEBNext Ultra II DNA Library Prep Kit for Illumina, New England Biolabs) including library size selection (SparQ beads, Qiagen) and quantification (Kapa Illumina GA with Revised Primers-SYBR Fast Universal Kit, Kapa Biosystems), and average fragment size determination (LabChip GX Touch Nucleic Acid Analyzer, PerkinElmer). Sequencing was performed on an Illumina NovaSeq 6000, generating 151 base pair paired-end reads. The samples yielded raw sequencing reads ranging from 38,474,441 to 133,152,604 reads.

For taxonomic classification, sequences were trimmed of adapters and subjected to quality control using *fastp* (v0.23.2) (6), aligned to the PhiX bacteriophage (GenBank accession NC_001422.1) using *Bowtie2* (v2.5.1) (7) with argument `--un-conc-gz` to retain non-PhiX sequences, then classified against bacterial genomes within the curated BeeRoLaMa database (8) using *Kraken 2* (v2.1.2) (9) with argument `--paired`. Default parameters were used except where otherwise noted. Classified counts confirmed the

Editor Frank J. Stewart, Montana State University, Bozeman, Montana, USA

Address correspondence to R. Ortega Polo, Rodrigo.OrtegaPolo@AGR.GC.CA, or M. M. Guarna, marta.guarna@outlook.com.

L. Tran and L. Lansing contributed equally to this article. The order of the first authors name is based on their contribution type.

The authors declare no conflict of interest.

See the funding table on p. 3.

Received 13 August 2024

Accepted 29 December 2024

Published 4 February 2025

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TABLE 1 Agricultural crops, sampling year, and province – region from which the experimental bee colonies were sampled (4, 5)^a

Crop	Years	Crop sample prefix	Province – region
Apple	2021	APP	Québec – Montérégie
Corn	2020	COR	Southwestern Ontario
Cranberry	2020, 2021	CRA	British Columbia – Fraser Valley, Québec – Centre-du-Québec
Canola oil	2020, 2021	CAC	Northern Alberta Southern Alberta Southern Manitoba
Lowbush blueberry	2021	LBB	Québec Saguenay – Lac-Saint-Jean Region
Soybean	2020	SOY	Southern Manitoba
Canola seed	2020, 2021	CAS	Southern Alberta
Highbush blueberry	2020, 2021	HBB	British Columbia – Fraser Valley

^aOur metadata includes GPS locations and sample names encoded with experimental details; a three-letter code specifying the agricultural crop (as per the 'crop sample prefix' column) followed by a two-digit site number (from 01 to 05 for 2020 and from 06 to 10 for 2021) and ending with an "e" or "u" indicating whether the sampled bees were near (e = exposed) or far (u = unexposed) from the named crop. In the case of NCBI BioProject PRJNA1124990, the time point (T1–T4) is also indicated as part of the sample name.

expected core honey bee gut microbiome taxa as well as the occurrence of pathogens *Paenibacillus* spp., *Spiroplasma* spp., and *Melissococcus plutonius*.

Our data sets should prove useful for answering diverse biological and environmental questions regarding differences in microbial diversity and relative abundance across Canada over 2 years. Some of the pathogen, chemical, and pollen diversity data associated with these colonies have been reported (4, 5, 10–12) and can be used to study interactions between these factors and the bee gut microbiome composition. The availability of the metagenomic data sets announced here will also facilitate association analyses between microbiome composition and environmental conditions, and inform the use of honey bees and their microbiomes for environmental monitoring (2).

ACKNOWLEDGMENTS

This work was funded by the Government of Canada through Agriculture and Agri-Food Canada (AAFC) Genomics Research and Development Initiative (GRDI) funding (AAFC J-002368), Genome Canada (LSARP #16420), and the Ontario Genomics Institute (OGI-185). We would like to thank the team at the Centre d'expertise et des services Génome Québec for their timely communication and sequencing services, the support team for the AAFC Biocluster where the bioinformatic processing of the sequencing data was performed, the landowners and beekeepers who facilitated hive placements and sample collections, and the BeeCSI Consortium members (<https://beecsi.ca/beecsi-consortium/>), in particular those who participated in the collection of bee samples used for microbiome analysis.

AUTHOR AFFILIATIONS

¹Agriculture and Agri-Food Canada, Beaverlodge Research Farm, Beaverlodge, Alberta, Canada

²Agriculture and Agri-Food Canada, Lethbridge Research and Development Centre, Lethbridge, Alberta, Canada

³Department of Computer Science, University of Victoria, Victoria, British Columbia, Canada

⁴Department of Biology, York University, Toronto, Ontario, Canada

⁵School of Environmental Sciences, University of Guelph, Guelph, Ontario, Canada

⁶Department of Integrative Biology, University of Texas, Austin, Texas, USA

⁷Department of Biological Sciences, University of Lethbridge, Lethbridge, Alberta, Canada

⁸Department of Entomology, University of Manitoba, Winnipeg, Manitoba, Canada

⁹Département de biologie, Université Laval, Québec City, Québec, Canada

¹⁰Department of Biochemistry and Molecular Biology, University of British Columbia, Vancouver, British Columbia, Canada

PRESENT ADDRESS

E. M. Walsh, U.S. Department of Agriculture, Agricultural Research Service, Baton Rouge, Louisiana, USA

H. Jabbari, Department of Biomedical Engineering, University of Alberta, Edmonton, Canada

M. M. Guarna, Project Apis m, Salt Lake, Utah, USA

M. M. Guarna, Department of Biochemistry and Molecular Biology, University of British Columbia, Vancouver, British Columbia, Canada

AUTHOR ORCID*s*

L. Tran  <http://orcid.org/0009-0005-9130-6535>

L. Lansing  <http://orcid.org/0000-0001-9945-9097>

N. Morfin  <http://orcid.org/0000-0002-1841-2084>

N. Moran  <http://orcid.org/0000-0003-2983-9769>

S. E. Hoover  <http://orcid.org/0000-0001-9462-7039>

S. F. Pernal  <http://orcid.org/0000-0002-0290-4391>

R. W. Currie  <http://orcid.org/0000-0003-1146-900X>

P. Giovenazzo  <http://orcid.org/0000-0002-8080-4935>

E. Guzman-Novoa  <http://orcid.org/0000-0001-5632-0642>

H. Jabbari  <http://orcid.org/0000-0002-7155-2297>

L. J. Foster  <http://orcid.org/0000-0001-8551-4817>

A. Zayed  <http://orcid.org/0000-0003-3233-4585>

R. Ortega Polo  <http://orcid.org/0000-0003-3411-923X>

M. M. Guarna  <http://orcid.org/0000-0002-4638-9475>

FUNDING

Funder	Grant(s)	Author(s)
Canadian Government Agriculture and Agri-Food Canada (AAFC)		S. F. Pernal
		R. Ortega Polo
		M. M. Guarna
Genome Canada (GC)		S. E. Hoover
		R. W. Currie
		P. Giovenazzo
		E. Guzman-Novoa
		L. J. Foster
		A. Zayed

DATA AVAILABILITY

The raw metagenome sequences for all T2 samples are available as BioProject [PRJNA999720](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA999720) in the NCBI BioProject database (<https://www.ncbi.nlm.nih.gov/bioproject/>), and sequences for all time points for the HBB experiment are available as NCBI BioProject [PRJNA1124990](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1124990). Both BioProjects fall under the umbrella BioProject [PRJNA1033630](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1033630).

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